From tea to treatment; epigallocatechin gallate and its potential involvement in minimizing the metabolic changes in cancer

Tauber, Amanda L.; Schweiker, Stephanie S.; Levonis, Stephan M.

Published in:
Nutrition Research

DOI:
[10.1016/j.nutres.2019.12.004](https://doi.org/10.1016/j.nutres.2019.12.004)

Licence:
CC BY-NC-ND

[Link to output in Bond University research repository.](#)

Recommended citation(APA):
Tauber, A. L., Schweiker, S. S., & Levonis, S. M. (2020). From tea to treatment; epigallocatechin gallate and its potential involvement in minimizing the metabolic changes in cancer. *Nutrition Research*, 74, 23-36.
<https://doi.org/10.1016/j.nutres.2019.12.004>

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

For more information, or if you believe that this document breaches copyright, please contact the Bond University research repository coordinator.

Highlights

- Catechins in green tea are associated with slowing the proliferation of prostate cancer cells
- Epigallocatechin gallate, is the most bioactive catechin in green tea
- Epigallocatechin gallate's activity may result from influence over the PI3K/Akt/mTOR pathway
- Many prostate cancer tumours show a dysregulation of the PI3K/Akt/mTOR pathway
- Combating PI3K/Akt/mTOR hyperactivation may be a strategy to reduce prostate cancer aggression

**From Tea to Treatment; Epigallocatechin Gallate and its Potential Involvement
in Minimizing the Metabolic Changes in Cancer.**

Amanda L. Tauber^{1,2}, Stephanie S. Schweiker^{1,3}, and Stephan M. Levonis^{1,4}.

¹Faculty of Health Sciences and Medicine, Bond University, Gold Coast, 4226,
Australia.

² atauber@bond.edu.au

³ sschweiker@bond.edu.au

⁴ slevonis@bond.edu.au

Author Information

Corresponding Author

Email: slevonis@bond.edu.au

ph. +61 7 5595 4417

17 **List of Abbreviations**

18

19 ADT; androgen deprivation therapy

20 Akt; protein kinase B

21 ATP; adenosine triphosphate

22 C; (+)-catechin

23 CDK; cyclin-dependent kinase

24 CG; (-)-catechin gallate

25 CLL; chronic lymphatic leukemia

26 CRPCa; castration-resistant prostate cancer

27 CRTC2; CREB regulated transcription coactivator 2

28 EC; (-)-epicatechin

29 ECG; (-)-epicatechin gallate

30 EGC; (-)-epigallocatechin

31 EGCG; (-)-epigallocatechin gallate

32 GC; (+)-gallocatechin

33 GCG; (-)-gallocatechin gallate

34 K_i ; inhibition constant

35 mTOR; mammalian target of rapamycin

36 NF- κ B; nuclear factor kappa-light-chain-enhancer of activated B cells

37 nM; nanomolar

38 p85/p110kin; phosphatidylinositol-4,5-bisphosphate 3-kinase

39 PCa; prostate cancer

40 PI3K; phosphatidylinositol 3-kinase

41 Rb; retinoblastoma protein

- 42 ROS; reactive oxygen species
- 43 RTK; receptor tyrosine kinase
- 44 S6K1; ribosomal protein S6 kinase beta-1
- 45 TCM; traditional Chinese medicine

Abstract

As the most abundant bioactive polyphenol in green tea, epigallocatechin gallate (EGCG) is a promising natural product that should be utilized in the discovery and development of potential drug leads. Due to its association with chemoprevention, EGCG may find a role in the development of therapeutics for prostate cancer. Natural products have long been employed as a scaffold for drug design, as their already noted bioactivity can help accelerate the development of novel treatments. Green tea and the EGCG contained within have become associated with chemoprevention, and both *in vitro* and *in vivo* studies have correlated EGCG to inhibiting cell growth and increasing the metabolic stress of cancer cells, possibly giving merit to its long utilized therapeutic use in traditional therapies. There is accumulating evidence to suggest that EGCG's role as an inhibitor of the PI3K/Akt/mTOR signaling cascade, acting upon major axis points within cancer survival pathways. The purpose of this review is to examine the research conducted on tea along with EGCG in the areas of the treatment of and/or prevention of cancer. This review discusses *Camellia sinensis*, as well as the bioactive phytochemical compounds contained within. Clinical uses of tea are explored, and possible pathways for activity are discussed before examining the evidence for EGCG's potential for acting on these processes. EGCG is identified as being a possible lead phytochemical for future drug design investigations.

EGCG; Cancer; PI3K/Akt/mTOR; Prostate Cancer; Natural Products

1.0 Introduction

There are multiple strategies when it comes to drug design, including *de novo* design, structure-based, target-based screening, pharmacophore searching; however, one of the longest standing approaches is the study of natural products. Isolation of bioactive components within natural products can lead to a stand-alone treatment or present a structural basis for a more efficient novel drug design. Green tea is one such natural product and has been traditionally administered for therapeutic use. Large cohort studies have hinted at a positive correlation between green tea consumption and cancer chemoprevention in men diagnosed with prostate cancer [1-6]. It has been discovered that one of green tea's most prominent bioactive component, the flavanol (-)-epigallocatechin gallate (EGCG), is likely the source of this activity [3, 7-12]. EGCG has been noted to influence key enzymes in the PI3K/Akt/mTOR pathway, which is commonly dysregulated in the development of prostate cancer, and could potentially act in a similar form to the synthetic inhibitors being developed against this pathway [13]. Despite its potential, work still needs to be done to identify whether green tea or EGCG can be recommended as a chemopreventative.[13]. Much of the large cohort evidence struggles to differentiate between the effect of consuming green tea and lifestyle choices [14]. Thus although there are multiple claims and evidence to suggest the benefit of EGCG for cancer therapy, more research is needed in both the therapeutic mechanisms of actions and the clinical benefit. By reviewing literature in these areas, the relevance of this natural product may be brought to light.

2.0 Approach

This study utilized a series of medical databases, inclusive of PubMed, EMBASE, MedLine, and SciFinder for articles published in the past 20 years to obtain a viable and comprehensive depiction of our current understanding of EGCG and its potential involvement in minimizing the deregulated of biochemical pathways observed in cancers. Search strategies began with using keywords such as “epigallocatechin gallate” AND “prostate cancer”, or more generally “green tea” AND “metastasis”. As data were collected and the interacting pathways better comprehended, the search requests expanded to more expansively investigate the involved PI3K/Akt/mTOR pathway, history, and previous association of green tea as a chemopreventive medicine, and studies investigating the modern approach to targeting the metabolic pathways of cancer.

3. Green Tea. Source and Bioactivity.

3.1 Botanical Source

EGCG is most abundantly found in green tea; however, it is also present in black and oolong teas, along with trace amounts found in miscellaneous fruit and vegetables [15]. All three of the major tea varieties including black, oolong, and green, are sourced from the *Camellia sinensis* plant, which grows globally in warm and humid climates [16]. China, Indonesia, Sri Lanka, and southern India have a year-round harvesting and growing season, whereas areas such as northern-eastern

India and northern China have a shorter season due to the greater seasonal variations [17]. The *Camellia Sinensis* is harvested by hand, with the “flush,” consisting of the top leaves connected to the bud and part of the stem making up the basis of tea [18]. It is during the processing of this flush where the black, green, and oolong tea varieties differ (Figure 1). For green tea, the flushes are withered and rolled, then either steamed or pan-roasted to inactivate the polyphenol oxidases within the plant [19]. From here, green tea is relatively stable during storage until seeping. This varies from the processing of black and oolong tea, as they lack the primary steaming step performed in green tea, and consequentially have a lower proportion of bioactive components in the final product [20-22].

3.2 Active Components in Green Tea

Amongst the wide variety of bioactive components in green tea, the polyphenols are the most abundant (Figure 2). Comprising around 40% of green tea’s dry mass, these compounds are colorless and water-soluble, contributing to the bitterness of the final product [20, 23]. Other compounds including the stimulatory methylxanthines, caffeine, theobromine, and theophylline are also present in tea, along with L-theanine, tannins, gallic acid, oxalic acid, pectin, fluoride, minerals and vitamins such as B1, B2, C, and E which can be found at varying concentrations, the most predominant category are the flavonoids [24, 25]. Characterized by their 2-phenylbenzopyran ring, variations in the C-ring saturation and oxidation status of flavonoids divide the classifications up into eight different groups, in which the flavan-3-ols are the most abundant (Figure 3).

Compared to the darker black and oolong teas which have most of their flavanols converted into their theaflavins and thearubigins counterparts during oxidation, green tea maintains a far higher proportion of the more bioactive flavanols (Table 1) [10, 26]. Such flavonoids include quercetin, kaempferol, and myricetin and the flavones apigenin and luteolin, with the largest class being the catechins [27]. By mass, epigallocatechin gallate (EGCG) is the most predominant (7–74 mg/g), followed by epicatechin gallate (ECG) (1–41 mg/g), epigallocatechin (EGC) (0–36.5 mg/g), epicatechin (EC) (0.1–9.5 mg/g) and catechin (C) (0–5.8 mg/g) [28–30] (Figure 4). However, depending on the variety, brand and location of harvest, these concentrations may vary [31].

3.3 Bioactive Role

Catechins are hydroxy and gallate substitutions of the flavan-3-ol structure, each with relative bioactive effects [32]. Chemotherapeutically, the galloylated catechins, GC, EGC, GCG, and EGCG are noted to possess the most chemotherapeutically active role [33–37]. The combined use of green tea catechins has been associated with antioxidant activity, chemoprevention, anti-viral, anti-inflammatory and anti-diabetic activity [38–41]. However, with the trihydroxyl groups at carbons 3', 4', and 5' on the B-ring, and a gallate moiety esterified at carbon 3' on the C-ring, EGCG presents with the greatest anti-proliferative and pro-apoptotic activity against cancer cells compared to the other catechins [3, 7–12].

Studies show extracting EGCG from tea is most effective at 80°C using a 50% v/v ethanol solution as this prevented epimerization of the catechin, however, if using

fresh leaves, then the extraction should use 75% v/v ethanol to compensate for the higher moisture content [42]. A later 2014 study investigating various extraction solvents, including ethanol, methanol, and water at different time intervals, concluded that a 40-minute extraction with ethanol maintained the greatest proportion of the catechins [43].

3.4 Clinical Uses

Records of the production and attributed health benefits of tea have dated back to the *Cha Jing (Tea Bible)* by the Lu Yu of the Tang Dynasty, often with a focus towards its anti-inflammatory action [44]. Such traditional Chinese medicines are still appreciated due to their theoretical approach and long-documented history [45]. Nowadays, many commercially available drugs derive inspiration from natural products, such as the chemotherapeutics topotecan and docetaxel which are synthetic alterations of the natural products camptothecin (*Camptothec acuminata*) and paclitaxel (*Taxus brevifolia*), along with Vincristine (*Catharanthus roseus*) which is a natural product (often synthetically generated) from the Madagascar periwinkle (Figure 5) [46]. EGCG's association with chemoprevention has prompted investment towards furthering its potential clinical application with two studies initiated in March 2018 to investigate its effect at minimizing the chemotherapeutic damage done in patients undergoing lung or breast cancer treatments [47, 48].

4. Prostate Cancer & Current Treatments

From 2015, 3300 deaths per year were attributed to prostate cancer, making it one of the leading causes of cancer-related mortality, accounting for 16% of Australia's male cancer expenditure [49, 50]. With the major risk factor being age, patients are left with few options to decrease their susceptibility towards the disease. Consequently, greater reliance is placed upon detection and treatment [51]. If surgery or radiotherapy fails to remove the cancer growth, androgen deprivation therapy (ADT), primarily bicalutamide (trade name Casodex), serves as the first-line chemotherapeutic [49]. However, this treatment is only palliative, acting to suppress the androgen driven growth in the early stages. Within 14-30 months, ADTs typically become redundant as the cells mutate into an androgen-independent state known as castration-resistant prostate cancer (CRPCa) [49]. Whether or not CRPCa is initially metastatic, 60% of men develop the metastatic disease within five years, with most developing it within three [52].

From here, docetaxel (trade name 'Taxotere') is the preferred chemotherapeutic, and it is associated with extremely high rates of chemoresistance and only extends the nine months' lifespan by an average three months [49, 53-55]. Since 2010, alternative treatments including immunology, cabazitaxel, enzalutamide, and abiraterone acetate have been trailed, extending the life expectancy by up to 5 months [23, 56]. However, these have been associated with a poorer quality of life than docetaxel. Some of the more common drug-based treatment options in Australia are tabulated below (Table 3).

4.1 Green tea and Prostate Cancer

The need for intervention, which can reduce the incidence of metastasis of Prostate Cancer (PCa) without severely hindering the quality of life, is going to be crucial to address this global health issue. In both Japanese and Chinese populations, there is a lower incidence of many cancers, including PCa, primarily attributed to their diet of soy, low fat, and high fiber as means of chemoprevention. Furthermore, their high intake of green tea has a strong positive correlation to chemoprevention [1-6]. Daily consumption of 10 or more cups a day is seen to increase the age of onset and decrease metastasis of a variety of cancers, including PCa [57-59]. Although not PCa, studies using squamous cell carcinomas cells show that the therapeutic index of 10 μ M can be reached with regular consumption of green tea [60]. Using a preparation known as polyphenol E, it was found that the maximum tolerable dose of green tea was 4.2 g/m² (equivalent to 20-30 cups of green tea) when tested on metachronous colorectal cells [61, 62].

The primary side effects, including polydipsia and urinary frequency, were suspected to be due to the caffeine content [57, 63-65]. However, with or without caffeine present, there was no significant difference in green tea's ability to inhibit angiogenesis *in vivo* [66]. Studies observing green tea's influence over PCa cell survival show a decrease in proliferation of androgen insensitive cells due to the bioactive components in green tea [6].

4.2 Bioavailability

The primary concern regarding the use of EGCG and other green tea preparations clinically was their low bioavailability [67]. The non-gallated green tea catechins undergo glucuronidation and sulfation *in vivo*, preventing their chemopreventive activity [68]. This observed in patients following a 6-week trial of oral green tea consumption, where 50% of EGCG in the prostate tissue appeared in its methylated form, consequentially decreasing the chemopreventive activity in the cells [69]. However, this may be combatted by the combined use of EGCG and quercetin (another polyphenol found in tea) [70]. Quercetin was seen to inhibit the catechol-O-methyltransferase and the multidrug-resistant proteins responsible for the methylation of EGCG and to improve EGCG's chemopreventive activity [70].

5.0 PI3K/AKT/MTOR PATHWAY IN CANCER

To better understand EGCG's potential clinical benefit, it is fitting to examine the likely intracellular signaling pathways affected by EGCG. It is unclear whether EGCG's activity is pro or antioxidant [36, 71, 72], much of EGCG's chemotherapeutic action is attributed to its influence over the PI3K/Akt/mTOR pathway. Defined by the key proteins; phosphatidylinositol 3-kinase (PI3K), protein kinase B (Akt), and mammalian target of rapamycin (mTOR), this pathway is a key regulator of metabolism, cell cycle, and preventing apoptosis; thus its hyperactivation is greatly involved in promoting the hallmarks of cancer [37, 73]. Many various carcinomas and prostate cancers observe the dysregulation of the PI3K/Akt/mTOR pathway, and this mutation is often a characteristic of chemoresistant cancer types [74].

269

270 This pathway is typically activated in response to the binding of hormonal or
271 mitogenic ligands to a receptor tyrosine kinase, phosphorylating the intracellular
272 subunit and activating the p85 and p110 kinase receptor units of the PI3K
273 heterodimer [75]. Activation induces the addition of ATP to PIP₂ in the cell
274 membrane, forming the PIP₃ signaling molecule, which in turn activates the PH
275 subunit of Akt, recruiting the protein to the cell membrane (Figure 6). This is followed
276 by the phosphorylation of phosphoinositide-dependent protein kinase-1 at the T308
277 residue, activating the complex [76]. Akt goes onto activate a number of intracellular
278 signaling processes, each holding influencing cell survival, proliferation, and growth,
279 primarily mediated through the two mTOR multiprotein complexes mTORC1 and
280 mTORC2 [77, 78].

281

282 **5.1 Pro survival and acceleration of growth**

283

284 Pro-survival and growth signaling are vital to cancer progression. With control over
285 the cell cycle and suppression of apoptosis, overactivation of the PI3K/Akt/mTOR
286 pathway can promote cancer cell survival [79]. The activated Akt phosphorylates and
287 inactivates glycogen synthase kinase-3 beta, preventing the activation of tumor
288 suppressor p53 and degradation of cyclin D [80]. Akt also induces the degradation of
289 p53 by inhibiting the transduction of nuclear-localized E3 ubiquitin ligase [81]. Now
290 remaining active in the nucleus, CD1 binds and activates the cyclin-dependent
291 kinase (CDK) proteins [82]. CDK4 and CDK6 inhibit the tumor suppressor
292 retinoblastoma protein (Rb), preventing the inhibitor of transcription factors G2F and
293 enabling the progression from G₁ to S-phase [82]. This amplified through mTORC1's

activation of ribosomal protein S6 kinase beta-1, F-box only protein 4, and inhibition of N-eukaryotic initiator factor, thus enhancing the stability of genes involved in S-phase entry [83]. Akt further ensures the activity of CDKs by inhibiting p21^{Cip1/Waf} and p27^{Kip1} [84, 85]. Direct phosphorylation of p21 also inhibits proliferating cell nuclear antigen, a suppressor of DNA replication [86]. Thus, in a multifactorial mechanism, the activation of the Akt pathway promotes and protects the progression through the cell cycle. Studies have observed that the cell cycle can be arrested at the G₁ phase through the inhibition of PI3K [87].

Another vital aspect of cell survival is the prevention of apoptosis. Through the breakdown of forkhead box O3 (FOXO3), Akt prevents the activation of p27, p21 p15, and p19, along with other proapoptotic genes such as BH3-only protein, Fas ligand and the p53 upregulated modulator of apoptosis [88]. Studies of primary chronic lymphatic leukemia B cells show that the constitutively active Akt increases expression and stability of the induced myeloid leukemia cell differentiation protein Mcl-1, the X-linked inhibitor of apoptosis protein and antiapoptotic B-cell lymphoma-extra large proteins, thus contributing to the inhibition of apoptosis & extending the longevity of diseased cells [78]. Furthermore, the cytochrome C induced apoptotic signaling pathway is inhibited by Akt at caspase 9, thereby promoting cell survival during cellular stress typical of the cancer environment.

5.2 Glycolysis

Another key hallmark of cancer is the switch towards anaerobic metabolism, described as the Warburg Effect [89]. This avoids the reliance on oxygen for energy production and is often correlated with tumor aggressiveness as it equips the cells with a rapid source of energy and intermediates for growth [90-93]. Targeting enzymes that promote the Warburg effect, such as the PI3K/Akt/mTOR pathway, act as a promising strategy to target the metabolic adaptations of cancer cells [94-97].

By phosphorylating the AS160 substrate on the glucose transporter type 4 receptor, AKT prompts its translocation to the cell surface and increases the cell's intake of glucose [98]. Downstream from the mTOR axis, there is the activation of other metabolism modulations that act to promote energy production and consumption within the cell [99]. PI3K phosphorylation and inhibition of FOXO1 and (downstream) CREB regulated transcription coactivator 2 inhibiting the fasting regulation of gluconeogenesis [100]. Storage of this excess glycogen is promoted through the inhibition of the glycogen synthase GSK3 β , which indirectly lowers the levels of c-Myc (noted in leukemic blast cells) [101]. Overall this prevents the inhibition of the hypoxic induced gene, promoting the Warburg Effect [102].

6.0 Targeting PI3K/Akt/mTOR

A genetic study of 218 prostate cancer tumors showed that 42% of primary growths and 100% of metastasizes displayed a genomic dysregulation of the PI3K/Akt/mTOR pathway [13], thus targeting key axis points within this pathway might be vital in

reducing the aggressiveness CRPCa. A meta-analysis in 2018 suggested that PI3K/Akt/mTOR pathway inhibitors can significantly improve the survival of patients with advanced solid tumors [74]. With a range of synthetic inhibitors being proposed, including dual mTORC1/2, dual PI3K/mTOR, Pan-PI3K, isoform-specific PI3K, and second-generation Akt inhibitors (Table 4), the therapeutic demand for a range of PI3K/Akt/mTOR inhibitors is evident.

6.1 Evidence for EGCGs role

EGCG itself acts as a competitive inhibitor (K_i 380 nM) of the common class 1 isomers of PI3K ($PI3K\alpha$, $PI3K\beta$, $PI3K\gamma$, and $PI3K\delta$), preventing the initial phosphorylation of Akt [103-108]. The binding mode of EGCG is noted to be similar to the PI3K inhibitor LY294002 [37]. It should be noted that within the LNCaP and PC-3 PCa cell lines, EGCG had no significant effect on the phosphorylation of PI3K at the Ser437 residue. The lack of the phosphatase and tensin homolog allele was suspected to cause the non-response [37]. EGCG also inhibits mTOR (K_i of 320 ± 24 nM [37], aligning itself in a similar category to the synthetic dual PI3K/Akt/mTOR inhibitors. These non-selective inhibitors display more promising effects as both pre-clinically and clinically, they are better equipped at overcoming the compensatory feedback mechanisms [37, 103-105].

Including its activity against the central PI3K/Akt/mTOR axis points, EGCG also interferes with the signaling cascade downstream from mTOR to reactivate the apoptotic signaling. Similar to other chemopreventive natural products such as curcumin, caffeinic acid, and capsaicin, EGCG is inhibitory against the transcription

factor; nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) [109]. NF- κ B is redox-responsive, and is highly implicated with the cancer cell proliferation and survival [110-112]. Although not specifically in PCa, when the squamous cell carcinomas cell line, A431 was treated with doses of 30-80 μ M of EGCG, the EGCG was seen to suppress the activation of NF- κ B, to which there is substantial evidence of crosstalk between the two pathways [113] [114]. Depleting the levels of NF- κ B in both the nucleus and cytoplasm, cancer cells were no longer protected against apoptosis, resulting in cell death [115]. Comparing the responsive dose of EGCG required to inhibit NF- κ B displayed an evident selectivity towards the cancerous A431 cell line over the non-cancerous normal human epidermal keratinocytes [109]. Furthermore, with the correlation between NF- κ B expression and PCa resistance against Docetaxel, the inhibition of NF- κ B (via BAY11-7082 inhibitor), appeared to reverse this resistance, and maybe the key to improving the efficacy of PCa drugs [116].

As a catechin, EGCG has a single-electron reduction potential enabling it to act as a scavenger for reactive oxygen species (ROS), and its pro-oxidant nature strongly contributes to pro-apoptotic activity. EGCG is susceptible to oxidation by H₂O₂ [36, 117, 118]. The then oxidized EGCG forms a cytotoxic o-quinone, which later reacts with glutathione to form various ROS [118, 119]. These ROS are suspected to downregulate Bcl-2 and Mcl-1 [36, 120]. When EGCG is administered in combination with arsenic trioxide (Trisenox®), a natural product based chemotherapeutic used in acute promyelocytic leukemia, the production of ROS was greater than seen with either used alone [36]. EGCG also displays some selectivity to cancerous cells, with apoptosis induced in the cancerous A431 cell line, but not normal epidermal

keratinocyte counterparts [109]. Thus, the combined increase in ROS and depletion of NF- κ B, EGCG is seen to counteract the prosurvival signaling enacted by PI3K/Akt/mTOR hyperactivation.

7.0 Discussion and Conclusion

There a deficit in our collective knowledge in the area of EGCG's role in the occurrence and treatment of cancer, as well as that of tea products in this same area, and this is indicative of the future work that might be done to address this. Due to EGCG being considered a pan assay interference compound, it can be assumed that other pathways are affected beyond PI3K/Akt/mTOR since its structural properties are conducive to broad interactions [121, 122]. Thus, there is a concern with its use to guide synthetic drug design. EGCG also has poor stability, it rapidly oxidizes in solution, and is rapidly metabolized in vivo [123-125]. Even so, the evidence provided in laboratory and clinical studies gives encouraging support for the further investigation of this phytochemical and its botanical source. Further study may take the form of clinical trials to assess the use of EGCG or tea products as adjunct natural therapies alongside traditional chemotherapy, or another promising area of work may be computational analysis for guided drug design, with EGCG as a lead compound. Although there is the potential for nonspecific interactions of such compounds when examined via in vitro assay, in vivo evidence encourages research to continue in this area.

Overcoming the metabolic adaptations of metastatic prostate cancer continues to be a major hurdle in producing effective treatments without severely hindering the

patient's quality of life. Due to the multifactorial nature of many cancers, in particular, CRPCa, single-target drugs are often redundant due to crosstalk within the prosurvival cascades, such as the PI3K/Akt/mTOR pathway. However, natural product EGCG may hold the solution. Acknowledged for its tolerability and chemotherapeutic activity against a variety of cancers, EGCG acts upon a range of targets within the PI3K/Akt/mTOR cascade to promote the selective apoptosis of cancer cells. With a growing risk of many late-stage cancers, investigating tolerable options such as EGCG may be essential for cancer treatment going forward. Whether effective on its own or to be utilized as adjuvant therapy, EGCG shows potential as a chemopreventative or sensitizer and may have the potential to lead further synthetic drug design.

8.0 Acknowledgments

This work was supported by Bond University. The authors declare no conflicts of interest.

437 **9.0 References**

- 438 [1] Thakur VS, Gupta K, Gupta S. Green tea polyphenols causes cell cycle arrest
439 and apoptosis in prostate cancer cells by suppressing class I histone deacetylases.
440 Carcinogenesis. 2012;33:377-84.
- 441 [2] Bettuzzi S, Brausi M, Rizzi F, Castagnetti G, Peracchia G, Corti A.
442 Chemoprevention of human prostate cancer by oral administration of green tea
443 catechins in volunteers with high-grade prostate intraepithelial neoplasia: a
444 preliminary report from a one-year proof-of-principle study. Cancer research.
445 2006;66:1234-40.
- 446 [3] Mukhtar H, Ahmad N. Green tea in chemoprevention of cancer. Toxicological
447 Sciences. 1999;52:111-7.
- 448 [4] Bushman JL. Green tea and cancer in humans: a review of the literature. Nutrition
449 and Cancer. 1998;31:151-9.
- 450 [5] Gupta S, Ahmad N, Mukhtar H. Prostate cancer chemoprevention by green tea.
451 Seminars in Urologic Oncology. 1999;17:70-6.
- 452 [6] Gupta S, Ahmad N, Nieminen AL, Mukhtar H. Growth inhibition, cell-cycle
453 dysregulation, and induction of apoptosis by green tea constituent (-)-
454 epigallocatechin-3-gallate in androgen-sensitive and androgen-insensitive human
455 prostate carcinoma cells. Toxicology and Applied Pharmacology. 2000;164:82-90.
- 456 [7] Ju J, Lu G, Lambert JD, Yang CS. Inhibition of carcinogenesis by tea
457 constituents. Seminars in Cancer Biology. 2007;17:395-402.
- 458 [8] Yang C. Inhibition of carcinogenesis by tea. Nature. 1997;389:134-5.
- 459 [9] Lambert JD, Yang CS. Mechanisms of Cancer Prevention by Tea Constituents.
460 Journal of Nutrition. 2003;133:3262-7.

461 [10] Du GJ, Zhang Z, Wen XD, Yu C, Calway T, Yuan CS, et al. Epigallocatechin
 462 Gallate (EGCG) is the most effective cancer chemopreventive polyphenol in green
 463 tea. *Nutrients*. 2012;4:1679-91.

464 [11] Ahmad N, Feyes DK, Nieminen A-L, Agarwal R, Mukhtar H. Green Tea
 465 Constituent Epigallocatechin-3-Gallate and Induction of Apoptosis and Cell Cycle
 466 Arrest in Human Carcinoma Cells. *Journal of the National Cancer Institute*.
 467 1997;89:1881-6.

468 [12] Hou Z, Lambert JD, Chin KV, Yang CS. Effects of tea polyphenols on signal
 469 transduction pathways related to cancer chemoprevention. *Mutation Research*.
 470 2004;555:3-19.

471 [13] Taylor BS, Schultz N, Hieronymus H, Gopalan A, Xiao Y, Carver BS, et al.
 472 Integrative genomic profiling of human prostate cancer. *Cancer Cell*. 2010;18:11-22.

473 [14] Lin Y-w, Hu Z-h, Wang X, Mao Q-q, Qin J, Zheng X-y, et al. Tea consumption
 474 and prostate cancer: an updated meta-analysis. *World J Surg Oncol*. 2014;12:38-.

475 [15] Chacko SM, Thambi PT, Kuttan R, Nishigaki I. Beneficial effects of green tea: a
 476 literature review. *Chinese medicine*. 2010;5:13-.

477 [16] Bhagat R, Baruah RD, Safique S. Climate and tea [*Camellia sinensis* (L.) O.
 478 Kuntze] production with special reference to North Eastern India: a review. *Journal of*
 479 *Environmental Research And Development* Vol. 2010;4.

480 [17] Awasom I. Tea. *Journal of Agricultural & Food Information*. 2011;12:12-22.

481 [18] Botwright T. Growth and quality of green tea (*Camellia sinensis* var *sinensis*) /
 482 by Tina Botwright. 1997.

483 [19] Tounekti T, Joubert E, Hernández I, Munné-Bosch S. Improving the Polyphenol
 484 Content of Tea. *Critical Reviews in Plant Sciences*. 2013;32:192-215.

485 [20] Sajilata MG, Bajaj PR, Singhal RS. Tea Polyphenols as Nutraceuticals.
 486 Comprehensive Reviews in Food Science and Food Safety. 2008;7:229-54.

487 [21] Astill, Birch, Dacombe, Humphrey, Martin. Factors affecting the caffeine and
 488 polyphenol contents of black and green tea infusions. Journal of Agricultural and
 489 Food Chemistry. 2001;49:5340-7.

490 [22] Friedman M, Levin CE, Choi SH, Lee SU, Kozukue N. Changes in the
 491 Composition of Raw Tea Leaves from the Korean Yabukida Plant during
 492 High-Temperature Processing to Pan-Fried Kamairi-Cha Green Tea. Journal of Food
 493 Science. 2009;74:C406-C12.

494 [23] Agarwal N, Di Lorenzo G, Sonpavde G, Bellmunt J. New agents for prostate
 495 cancer. Annals of Oncology. 2014;25:1700-9.

496 [24] Anaya AL, Cruz-Ortega R, Waller GR. Metabolism and ecology of purine
 497 alkaloids. Frontiers in bioscience : a journal and virtual library. 2006;11:2354.

498 [25] Cloughley JB, Ellis RT, Harris N. Black tea manufacture. Annals of Applied
 499 Biology. 1981;99:367-74.

500 [26] Graham HN. Green tea composition, consumption, and polyphenol chemistry.
 501 Preventive Medicine. 1992;21:334-50.

502 [27] Panche AN, Diwan AD, Chandra SR. Flavonoids: an overview. Journal of
 503 nutritional science. 2016;5:e47.

504 [28] Friedman M, Levin CE, Choi SH, Kozukue E, Kozukue N. HPLC Analysis of
 505 Catechins, Theaflavins, and Alkaloids in Commercial Teas and Green Tea Dietary
 506 Supplements: Comparison of Water and 80% Ethanol/Water Extracts. Journal of
 507 Food Science. 2006;71:C328-C37.

508 [29] Khokhar S, Magnusdottir SGM. Total phenol, catechin, and caffeine contents of
509 teas commonly consumed in the United kingdom. *Journal of agricultural and food*
510 *chemistry*. 2002;50:565.

511 [30] Lee B-L, Ong C-N. Comparative analysis of tea catechins and theaflavins by
512 high-performance liquid chromatography and capillary electrophoresis. *Journal of*
513 *Chromatography A*. 2000;881:439-47.

514 [31] Chen Z, Zhu QY, Tsang D, Huang Y. Degradation of green tea catechins in tea
515 drinks. *Journal of agricultural and food chemistry*. 2001;49:477.

516 [32] Sáez-Ayala M, Fernández-Pérez MP, Chazarra S, McHedlishvili N, Tárraga-
517 Tomás A, Rodríguez-López JN. Factors Influencing the Antifolate Activity of
518 Synthetic Tea-Derived Catechins. *Molecules*. 2013;18:8319-41.

519 [33] Shimizu M, Fukutomi Y, Ninomiya M, Nagura K, Kato T, Araki H, et al. Green
520 tea extracts for the prevention of metachronous colorectal adenomas: a pilot study.
521 *Cancer Epidemiology, Biomarkers & Prevention*. 2008;17:3020-5.

522 [34] Fujiki, Suganuma, Imai, Nakachi. Green tea: Cancer preventive beverage and/or
523 drug. *Cancer Letters*. 2002;188:9-13.

524 [35] Tsao AS, Liu D, Martin J, Tang Xm, Lee JJ, El-Naggar AK, et al. Phase II
525 Randomized, Placebo-Controlled Trial of Green Tea Extract in Patients with High-
526 Risk Oral Premalignant Lesions. *Cancer Prevention Research*. 2009;2:931-41.

527 [36] Nakazato T, Ito K, Ikeda Y, Kizaki M. Green tea component, catechin, induces
528 apoptosis of human malignant B cells via production of reactive oxygen species.
529 *Clinical Cancer Research*. 2005;11:6040-9.

530 [37] Van Aller GS, Carson JD, Tang W, Peng H, Zhao L, Copeland RA, et al.
531 Epigallocatechin gallate (EGCG), a major component of green tea, is a dual

532 phosphoinositide-3-kinase/mTOR inhibitor. Biochemical and Biophysical Research
533 Communications. 2011;406:194-9.

534 [38] Khan N, Mukhtar H. Tea polyphenols for health promotion. Life Sciences.
535 2007;81:519-33.

536 [39] Lakenbrink C, Lapczynski S, Maiwald B, Engelhardt UH. Flavonoids and other
537 polyphenols in consumer brews of tea and other caffeinated beverages. Journal of
538 agricultural and food chemistry. 2000;48:2848.

539 [40] Zaveri NT. Green tea and its polyphenolic catechins: Medicinal uses in cancer
540 and noncancer applications. Life Sciences. 2006;78:2073-80.

541 [41] Henry JP, Stephens-Larson P. Reduction of chronic psychosocial hypertension
542 in mice by decaffeinated tea. Hypertension (Dallas, Tex : 1979). 1984;6:437.

543 [42] Liang H, Liang Y, Dong J, Lu J. Tea extraction methods in relation to control of
544 epimerization of tea catechins. Journal of the Science of Food and Agriculture.
545 2007;87:1748-52.

546 [43] Ahmad RS, Butt MS, Huma N, Sultan MT, Arshad MU, Mushtaq Z, et al.
547 Quantitative and Qualitative Portray of Green Tea Catechins (Gtc) Through Hplc.
548 International Journal of Food Properties. 2013;17.

549 [44] Yang CS, Chen G, Wu Q. Recent scientific studies of a traditional chinese
550 medicine, tea, on prevention of chronic diseases. Journal of traditional and
551 complementary medicine. 2014;4:17-23.

552 [45] Keji C, Hao XU. The integration of traditional Chinese medicine and Western
553 medicine. European Review. 2003;11:225-35.

554 [46] Demain AL, Vaishnav P. Natural products for cancer chemotherapy. Microbial
555 biotechnology. 2011;4:687-99.

556 [47] Zhao HX. Study of Epigallocatechin-3-gallate (EGCG) for Esophagus Protection
557 in Patients With Lung Cancer Receiving Radial Radiotherapy. In: Institute NC, editor.
558 US: U.S. National Library of Medicine; 2019.

559 [48] Zhao HX. Study of Epigallocatechin-3-gallate (EGCG) for Skin Prevention in
560 Patients With Breast Cancer Receiving Adjuvant Radiotherapy. In: Institute NC,
561 editor. US: U.S. National Library of Medicine; 2018.

562 [49] Bachmann SB, Frommel SC, Camicia R, Winkler HC, Santoro R, Hassa PO.
563 DTX3L and ARTD9 inhibit IRF1 expression and mediate in cooperation with ARTD8
564 survival and proliferation of metastatic prostate cancer cells. *Molecular Cancer*.
565 2014;13.

566 [50] AIHW. Cancer in Australia 2017. *Cancer*. Canberra: Australian Institute of
567 Health and Welfare; 2017.

568 [51] Damber J-E, Aus G. Prostate cancer. *The Lancet*. 2008;371:1710-21.

569 [52] Moreira DM, Howard LE, Sourbeer KN, Amarasekara HS, Chow LC, Cockrell
570 DC, et al. Predictors of Time to Metastasis in Castration-resistant Prostate Cancer.
571 *Urology*. 2016;96:171-6.

572 [53] Lorenzo G, Buonerba C, Autorino R, Placido S, Sternberg C. Castration-
573 Resistant Prostate Cancer. *Drugs*. 2010;70:983-1000.

574 [54] Kirby M, Hirst C, Crawford ED. Characterising the castration-resistant prostate
575 cancer population: a systematic review. *International Journal of Clinical Practice*.
576 2011;65:1180-92.

577 [55] Petrylak DP, Tangen CM, Hussain MHA, Lara PN, Jones JA, Taplin ME, et al.
578 Docetaxel and Estramustine Compared with Mitoxantrone and Prednisone for
579 Advanced Refractory Prostate Cancer. *The New England Journal of Medicine*.
580 2004;351:1513-20.

581 [56] Ezzell EE, Chang KS, George BJ. New agents in the arsenal to fight castrate-
582 resistant prostate cancer. *Current Oncology Reports*. 2013;15:239-48.

583 [57] Lu G, Liao J, Yang G, Reuhl KR, Hao X, Yang CS. Inhibition of adenoma
584 progression to adenocarcinoma in a 4-(methylnitrosamino)-1-(3-pyridyl)-1-butanone-
585 induced lung tumorigenesis model in A/J mice by tea polyphenols and caffeine.
586 *Cancer research*. 2006;66:11494-501.

587 [58] Kono S, Ikeda M, Tokudome S, Kuratsune M. A Case-Control Study of Gastric
588 Cancer and Diet in Northern Kyushu, Japan. *Japanese Journal of Cancer Research*.
589 1988;79:1067-74.

590 [59] Nakachi K, Matsuyama S, Miyake S, Suganuma M, Imai K. Preventive effects of
591 drinking green tea on cancer and cardiovascular disease: Epidemiological evidence
592 for multiple targeting prevention. *BioFactors*. 2000;13:49-54.

593 [60] Reygaert W. An Update on the Health Benefits of Green Tea. *Beverages*.
594 2017;3.

595 [61] Pisters K, Newman R, Coldman B, Shin D, Khuri F, Hong W, et al. Phase I trial
596 of oral green tea extract in adult patients with solid tumors. *Journal of Clinical*
597 *Oncology*. 2001;19 1830-8.

598 [62] Fujiki H, Suganuma M. Green tea: an effective synergist with anticancer drugs
599 for tertiary cancer prevention. *Cancer Letters*. 2012;324:119-25.

600 [63] Jung Y, Ellis L. Inhibition of tumour invasion and angiogenesis by
601 epigallocatechin gallate (EGCG), a major component of green tea. *International*
602 *Journal of Experimental Pathology*. 2001;82:309-16.

603 [64] Bertolini F, Fusetti L, Rabascio C, Cinieri S, Martinelli G, Pruneri G. Inhibition of
604 angiogenesis and induction of endothelial and tumor cell apoptosis by green tea in

605 animal models of human high-grade non-Hodgkin's lymphoma. *Leukemia*.
 606 2000;14:1477-82.
 607 [65] Lai HC, Chao WT, Chen YT, Yang VC. Effect of EGCG, a major component of
 608 green tea, on the expression of Ets-1, c-Fos, and c-Jun during angiogenesis in vitro.
 609 *Cancer Letters*. 2004;213:181-8.
 610 [66] Kanwar J, Taskeen M, Mohammad I, Huo C, Chan TH, Dou QP. Recent
 611 advances on tea polyphenols. *Frontiers in Bioscience - Elite*. 2012;E4:111-31.
 612 [67] Chu C, Deng J, Man Y, Qu Y. Green Tea Extracts Epigallocatechin-3-gallate for
 613 Different Treatments. *Biomed Res Int*. 2017;2017:5615647-.
 614 [68] Lu H, Meng X, Yang CS. Enzymology of methylation of tea catechins and
 615 inhibition of catechol-O-methyltransferase by (-)-epigallocatechin gallate. *Drug*
 616 *Metabolism and Disposition*. 2003;31:572-9.
 617 [69] Wang P, Aronson WJ, Huang M, Zhang Y, Lee R-P, Heber D, et al. Green tea
 618 polyphenols and metabolites in prostatectomy tissue: implications for cancer
 619 prevention. *Cancer Prev Res (Phila)*. 2010;3:985-93.
 620 [70] Wang P, Heber D, Henning SM. Quercetin increased bioavailability and
 621 decreased methylation of green tea polyphenols in vitro and in vivo. *Food & function*.
 622 2012;3:635-42.
 623 [71] Kumar N, Shibata D, Helm J, Malafa M, Coppola D. Green tea polyphenols in
 624 the prevention of colon cancer. *Frontiers in Bioscience*. 2007;12:2309-15.
 625 [72] Mak JC. Potential role of green tea catechins in various disease therapies:
 626 progress and promise. *Clinical and Experimental Pharmacology and Physiology*.
 627 2012;39:265-73.
 628 [73] Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*.
 629 2011;144:646-74.

630 [74] Li X, Dai D, Chen B, Tang H, Xie X, Wei W. Efficacy of PI3K/AKT/mTOR
631 pathway inhibitors for the treatment of advanced solid cancers: A literature-based
632 meta-analysis of 46 randomised control trials. PLoS One. 2018;13:e0192464.

633 [75] Liu P, Cheng H, Roberts TM, Zhao JJ. Targeting the phosphoinositide 3-kinase
634 pathway in cancer. Nature Reviews Drug Discovery. 2009;8:627-44.

635 [76] Alessi DR, James SR, Downes CP, Holmes AB, Gaffney PRJ, Reese CB, et al.
636 Characterization of a 3-phosphoinositide-dependent protein kinase which
637 phosphorylates and activates protein kinase B α . Current Biology. 1997;7:261-9.

638 [77] Guertin DA, Sabatini DM. Defining the role of mTOR in cancer. Cancer Cell.
639 2007;12:9-22.

640 [78] Longo PG, Laurenti L, Gobessi S, Sica S, Leone G, Efremov DG. The Akt/Mcl-1
641 pathway plays a prominent role in mediating antiapoptotic signals downstream of the
642 B-cell receptor in chronic lymphocytic leukemia B cells. Blood. 2008;111:846-55.

643 [79] Wegiel B, Bjartell A, Culig Z, Persson JL. Interleukin-6 activates PI3K/Akt
644 pathway and regulates cyclin A1 to promote prostate cancer cell survival.
645 International journal of cancer. 2008;122:1521.

646 [80] Nayak G, Cooper GM. p53 is a major component of the transcriptional and
647 apoptotic program regulated by PI3-kinase/Akt/GSK3 signaling. Cell Death and
648 Disease. 2012;3:e400.

649 [81] Mayo LD, Donner DB. A phosphatidylinositol 3-kinase/Akt pathway promotes
650 translocation of Mdm2 from the cytoplasm to the nucleus. Proceedings of the
651 National Academy of Sciences of the United States of America. 2001;98:11598-603.

652 [82] Diehl JA, Cheng M, Roussel MF, Sherr CJ. Glycogen synthase kinase-3 β
653 regulates cyclin D1 proteolysis and subcellular localization. Genes & Development.
654 1998;12:3499-511.

655 [83] Fingar DC, Richardson CJ, Tee AR, Cheatham L, Tsou C, Blenis J. mTOR
656 Controls Cell Cycle Progression through Its Cell Growth Effectors S6K1 and 4E-
657 BP1/Eukaryotic Translation Initiation Factor 4E. *Molecular and Cellular Biology*.
658 2004;24:200-16.

659 [84] Zhou BP, Liao Y, Xia W, Spohn B, Lee M-H, Hung M-C. Cytoplasmic localization
660 of p21Cip1/WAF1 by Akt-induced phosphorylation in HER-2/neu-overexpressing
661 cells. *Nature Cell Biology*. 2001;3:245.

662 [85] Polyak K, Lee M-H, Erdjument-Bromage H, Koff A, Roberts JM, Tempst P, et al.
663 Cloning of p27 Kip1 , a cyclin-dependent kinase inhibitor and a potential mediator of
664 extracellular antimitogenic signals. *Cell*. 1994;78:59-66.

665 [86] Cayrol C, Knibiehler M, Ducommun B. p21 binding to PCNA causes G1 and G2
666 cell cycle arrest in p53-deficient cells. *Oncogene*. 1998;16:311.

667 [87] Collado M, Medema RH, Garcia-Cao I, Dubuisson ML, Barradas M, Glassford J,
668 et al. Inhibition of the phosphoinositide 3-kinase pathway induces a senescence-like
669 arrest mediated by p27Kip1. *The Journal of biological chemistry*. 2000;275:21960.

670 [88] Zhang X, Tang N, Hadden TJ, Rishi AK. Akt, FoxO and regulation of apoptosis.
671 *Biochimica et Biophysica Acta - Molecular Cell Research*. 2011;1813:1978-86.

672 [89] Cho SH, Ahna AK, Bhargavab P, Leeb C, Eischenc CM, McGuinnessd O, et al.
673 Glycolytic rate and lymphomagenesis depend on PARP14, an ADP
674 ribosyltransferase of the B aggressive lymphoma (BAL) family. *Proceedings of the*
675 *National Academy of Sciences*. 2011;108:15972-7.

676 [90] Iansante V, Choy PM, Fung SW, Liu Y, Chai JG, Dyson J, et al. PARP14
677 promotes the Warburg effect in hepatocellular carcinoma by inhibiting JNK1-
678 dependent PKM2 phosphorylation and activation. *Nature Communications*.
679 2015;6:7882.

680 [91] Iqbal MA, Gupta V, Gopinath P, Mazurek S, Bamezai RNK. Pyruvate kinase M2
681 and cancer: an updated assessment. National Centre of Applied Human Genetics,.
682 2014;588:2685-92.

683 [92] Warburg O, Posener K, Negelein E. On Metabolism of Tumors. Biochemische
684 Zeitschrift. 1924;152:319-44.

685 [93] Bluemlein K, Grüning N-M, Feichtinger RG, Lehrach H, Kofler B, Ralser M. No
686 evidence for a shift in pyruvate kinase PKM1 to PKM2 expression during
687 tumorigenesis. Oncotarget. 2011;2:393-400.

688 [94] Chen Z, Lu W, Garcia-Prieto C, Huang P. The Warburg effect and its cancer
689 therapeutic implications. Journal of Bioenergetics and Biomembranes. 2007;39:267-
690 74.

691 [95] Scatena R, Bottoni P, Pontoglio A, Giardina B. Revisiting the Warburg effect in
692 cancer cells with proteomics. The emergence of new approaches to diagnosis,
693 prognosis and therapy. Proteomics Clinical Applications 2010;4:143-58.

694 [96] Wang Z, Wang N, Chen J, Shen J. Emerging glycolysis targeting and drug
695 discovery from chinese medicine in cancer therapy. Evidence-Based
696 Complementary and Alternative Medicine. 2012;2012:873175.

697 [97] Schweiker SS, Tauber AL, Sherry ME, Levonis SM. Poly(ADP-
698 ribose)polymerase, member 14 (PARP14) Inhibitors and Possible Strategies for
699 Designing Selective PARP14 Inhibitors. Mini-Reviews in Medicinal Chemistry. 2018.

700 [98] Sano H, Kane S, Sano E, Miinea CP, Asara JM, Lane WS, et al. Insulin-
701 stimulated phosphorylation of a Rab GTPase-activating protein regulates GLUT4
702 translocation. The Journal of Biological Chemistry. 2003;278:14599-602.

703 [99] Sciarretta S, Forte M, Frati G, Sadoshima J. New Insights Into the Role of
704 mTOR Signaling in the Cardiovascular System. Circ Res. 2018;122:489-505.

705 [100] Oh K-J, Han H-S, Kim M-J, Koo S-H. CREB and FoxO1: two transcription
706 factors for the regulation of hepatic gluconeogenesis. *BMB Reports*. 2013;46:567-74.

707 [101] Swords R, Schenk, T., Stengel, S., Sylvina, V., Petrie, K., Perez, A., . . . Zelent,
708 A. (). . Inhibition of the PI3K/AKT/mTOR Pathway Leads to Down-Regulation of c-
709 Myc and Overcomes Resistance to ATRA in Acute Myeloid Leukemia. *Blood*.
710 2015;126.

711 [102] Xiang S, Gu H, Jin L, Thorne RF, Zhang XD, Wu M. LncRNA IDH1-AS1 links
712 the functions of c-Myc and HIF1 α via IDH1 to regulate the Warburg effect.
713 *Proceedings of the National Academy of Sciences of the United States of America*.
714 2018;115:E1465.

715 [103] Chiarini F, Fala F, Tazzari PL, Ricci F, Astolfi A, Pession A, et al. Dual
716 inhibition of class IA phosphatidylinositol 3-kinase and mammalian target of
717 rapamycin as a new therapeutic option for T-cell acute lymphoblastic leukemia.
718 *Cancer research*. 2009;69:3520-8.

719 [104] Serra V, Markman B, Scaltriti M, Eichhorn PJ, Valero V, Guzman M, et al.
720 NVP-BEZ235, a dual PI3K/mTOR inhibitor, prevents PI3K signaling and inhibits the
721 growth of cancer cells with activating PI3K mutations. *Cancer research*.
722 2008;68:8022-30.

723 [105] Maira SM, Stauffer F, Brueggen J, Furet P, Schnell C, Fritsch C, et al.
724 Identification and characterization of NVP-BEZ235, a new orally available dual
725 phosphatidylinositol 3-kinase/mammalian target of rapamycin inhibitor with potent in
726 vivo antitumor activity. *Molecular Cancer Therapeutics*. 2008;7:1851-63.

727 [106] Qin J, Xie LP, Zheng XY, Wang YB, Bai Y, Shen HF, et al. A component of
728 green tea, (-)-epigallocatechin-3-gallate, promotes apoptosis in T24 human bladder

729 cancer cells via modulation of the PI3K/Akt pathway and Bcl-2 family proteins.
 730 Biochemical and Biophysical Research Communications. 2007;354:852-7.
 731 [107] Natsume H, Adachi S, Takai S, Tokuda H, Matsushima-Nishiwaki R,
 732 Minamitani C, et al. (-)-Epigallocatechin gallate attenuates the induction of HSP27
 733 stimulated by sphingosine 1-phosphate via suppression of phosphatidylinositol 3-
 734 kinase/Akt pathway in osteoblasts. International Journal of Molecular Medicine.
 735 2009;24:197-203.
 736 [108] Kim MJ, Kim HI, Chung J, Jeong TS, Park HR. (-)-Epigallocatechin-3-gallate
 737 (EGCG) increases the viability of serum-starved A549 cells through its effect on Akt.
 738 American Journal of Chinese Medicine. 2009;37:723-34.
 739 [109] Ahmad N, Gupta S, Mukhtar H. Green tea polyphenol epigallocatechin-3-
 740 gallate differentially modulates nuclear factor kappaB in cancer cells versus normal
 741 cells. Archives of Biochemistry and Biophysics. 2000;376:338-46.
 742 [110] Baeuerle P. Function and Activation of NF-Kappa B in the Immune System.
 743 Annual Review of Immunology. 1994;12:141-79.
 744 [111] Waddick, Uckun. Innovative treatment programs against cancer: Nuclear
 745 factor-kB (NF-kB) as a molecular target. Biochemical Pharmacology. 1999;57:9-17.
 746 [112] Huguet C, Crepieux P, Laudet V. Rel/NF-kB transcription factors and Ikb
 747 inhibitors: Evolution from a unique common ancestor. Oncogene. 1997;15:2965-74.
 748 [113] Dong Z, Ma W, Huang C, Yang C. Inhibition of tumor promoter-induced
 749 activator protein 1 activation and cell transformation by tea polyphenols, (-)-
 750 epigallocatechin gallate, and theaflavins. Cancer research. 1997;57:4414-9.
 751 [114] Ahmad A, Biersack B, Li Y, Kong D, Bao B, Schobert R, et al. Targeted
 752 Regulation of PI3K/Akt/mTOR/NF-kB Signaling by Indole Compounds and their

753 Derivatives: Mechanistic Details and Biological Implications for Cancer Therapy.
 754 Anti-cancer Agents in Medicinal Chemistry. 2013;13:1002-13.
 755 [115] Baichwal VR, Baeuerle PA. Apoptosis: Activate NF- κ B or die? Current Biology.
 756 1997;7:94-6.
 757 [116] O'Neill AJ, Prencipe M, Dowling C, Fan Y, Mulrane L, Gallagher WM, et al.
 758 Characterisation and manipulation of docetaxel resistant prostate cancer cell lines.
 759 Molecular Cancer. 2011;10:126.
 760 [117] Yang G-Y, Liao J, Li C, Chung J, Yurkow EJ, Ho C-T, et al. Effect of black and
 761 green tea polyphenols on c-jun phosphorylation and H₂O₂ production in transformed
 762 and non-transformed human bronchial cell lines: Possible mechanisms of cell growth
 763 inhibition and apoptosis induction. Carcinogenesis. 2000;21:2035-9.
 764 [118] Yang G, Liao J, Kim K, Yurkow E, Yang C. Inhibition of growth and induction of
 765 apoptosis in human cancer cell lines by tea polyphenols. Carcinogenesis.
 766 1998;19:611-6.
 767 [119] Moridani M, Scobie H, Salehi P, O'Brien P. Catechin metabolism: Glutathione
 768 conjugate formation catalyzed by tyrosinase, peroxidase, and cytochrome p450.
 769 Chemical Research in Toxicology. 2001;14 841-8.
 770 [120] Pei X, Dai Y, Grant S. Synergistic induction of oxidative injury and apoptosis in
 771 human multiple myeloma cells by the proteasome inhibitor bortezomib and histone
 772 deacetylase inhibitors. Clinical Cancer Research: An Official Journal of the American
 773 Association for Cancer Research. 2004;10:3839-52.
 774 [121] Baell JB. Feeling Nature's PAINS: Natural Products, Natural Product Drugs,
 775 and Pan Assay Interference Compounds (PAINS). Journal of Natural Products.
 776 2016;79:616-28.

777 [122] Singh BN, Shankar S, Srivastava RK. Green tea catechin, epigallocatechin-3-
778 gallate (EGCG): mechanisms, perspectives and clinical applications. *Biochemical*
779 *pharmacology*. 2011;82:1807-21.

780 [123] Feng WY. Metabolism of green tea catechins: an overview. *Current drug*
781 *metabolism*. 2006;7:755-809.

782 [124] Mochizuki M, Yamazaki S, Kano K, Ikeda T. Kinetic analysis and mechanistic
783 aspects of autoxidation of catechins. *Biochimica et biophysica acta*. 2002;1569:35-
784 44.

785 [125] Smith TJ. Green Tea Polyphenols in drug discovery - a success or failure?
786 *Expert Opin Drug Discov*. 2011;6:589-95.

787 [126] Nakachi K, Matsuyama S, Miyake S, Suganuma M, Imai K. Preventive effects
788 of drinking green tea on cancer and cardiovascular disease: Epidemiological
789 evidence for multiple targeting prevention. *BioFactors*. 2000;13:49-54.

790 [127] Du GJ, Wang CZ, Qi LW, Zhang ZY, Calway T, He TC, et al. The synergistic
791 apoptotic interaction of panaxadiol and epigallocatechin gallate in human colorectal
792 cancer cells. *Phytotherapy Research*. 2013;27:272-7.

793 [128] Hara Y. Influence of tea catechins on the digestive tract. *Journal of Cellular*
794 *Biochemistry*. 1997;67:52-8.

795 [129] Belguise K, Guo S, Sonenshein GE. Activation of FOXO3a by the green tea
796 polyphenol epigallocatechin-3-gallate induces estrogen receptor alpha expression
797 reversing invasive phenotype of breast cancer cells. *Cancer research*.
798 2007;67:5763-70.

799 [130] Szabo E, Riffe M, Steinberg S, Birrer M, Linnoila R. Altered cJUN expression:
800 An early event in human lung carcinogenesis. *Cancer research*. 1996;56:305-15.

801 [131] Wodrich W, Volm M. Overexpression of oncoproteins in non-small cell lung
802 carcinomas of smokers. *Lung Cancer*. 1994;10:397.

803 [132] Lukka H, Waldron T, Klotz L, Winquist E, Trachtenberg J, on behalf of the
804 Genitourinary Cancer Disease Site G, et al. Maximal androgen blockade for the
805 treatment of metastatic prostate cancer—a systematic review. *Current Oncology*.
806 2006;13:81-93.

807 [133] Klotz L, Boccon-Gibod L, Shore ND, Andreou C, Persson B-E, Cantor P, et al.
808 The efficacy and safety of degarelix: a 12-month, comparative, randomized, open-
809 label, parallel-group phase III study in patients with prostate cancer. *BJU*
810 *International*. 2008;102:1531-8.

811 [134] Schröder FH, Whelan P, de Reijke TM, Kurth KH, Pavone-Macaluso M,
812 Mattelaer J, et al. Metastatic Prostate Cancer Treated by Flutamide versus
813 Cyproterone Acetate. *European Urology*. 2004;45:457-64.

814 [135] Parker C, Heidenreich A, Nilsson S, Shore N. Current approaches to
815 incorporation of radium-223 in clinical practice. *Prostate Cancer and Prostatic*
816 *Diseases*. 2018;21:37-47.

817 [136] Anassi E, Ndefo UA. Sipuleucel-T (Provenge) Injection: The First
818 Immunotherapy Agent (Vaccine) For Hormone-Refractory Prostate Cancer.
819 *Pharmacy and Therapeutics*. 2011;36:197-202.

820 [137] Nuhn P, De Bono JS, Fizazi K, Freedland SJ, Grilli M, Kantoff PW, et al.
821 Update on Systemic Prostate Cancer Therapies: Management of Metastatic
822 Castration-resistant Prostate Cancer in the Era of Precision Oncology. *European*
823 *Urology*. 2019;75:88-99.

824 [138] Schellhammer PF, Sharifi R, Block NL, Soloway MS, Venner PM, Lynn
825 Patterson A, et al. Nical benefits of bicalutamide compared with flutamide in

826 combined androgen blockade for patients with advanced prostatic carcinoma: Final
827 report of a double-blind, randomized, multicenter trial. *Urology*. 1997;50:330-6.

828 [139] Loblaw DA, Virgo KS, Nam R, Somerfield MR, Ben-Josef E, Mendelson DS, et
829 al. Initial Hormonal Management of Androgen-Sensitive Metastatic, Recurrent, or
830 Progressive Prostate Cancer: 2007 Update of an American Society of Clinical
831 Oncology Practice Guideline. *Journal of Clinical Oncology*. 2007;25:1596-605.

832 [140] Crawford ED, Phillips JM. Six-month gonadotropin releasing hormone (GnRH)
833 agonist depots provide efficacy, safety, convenience, and comfort. *Cancer*
834 *Management and Research*. 2011;3:201-9.

835

836 10.0 Figures

837
838 **Figure 1: Generalized processing protocol for Green, Oolong and Black Tea. Primary**
839 **differences in the catechin content of each tea variety result from the variation in**
840 **treatment during the processing of the *Camellia sinensis* flush.**

845

846 **Figure 2: Key bioactive components of green tea, broadly categorized into flavonoids,**
847 **methylxanthines, vitamins and other.**

848

Flavone

Flavonol

Flavanone

Flavan-3-ol

Flavanonol

Anthocyanidin

Chalcone

Aurone

849

850

851

852

Figure 3: Names and structures of the 2-phenylbenzopyrans (flavonoids), in which flavan-3-ol is the most predominant

Flavones

Apigenin

Lutelin

Flavonoids

Quercetin

Myricetin

Kaempferol

Flavan-3-ol (Catechins)

(-)-Epicatechin (EC)

(+)-Catechin (C)

(-)-Epigallocatechin (EGC)

(+)-Gallocatechin (GC)

(-)-Epicatechin gallate (ECG)

(-)-Catechin gallate (CG)

(-)-Epigallocatechin gallate (EGCG)

(-)-Gallocatechin gallate (GCG)

853

854

855

Figure 4: Structures, names, and abbreviations of the major flavonols found in green tea

Topotecan (Hycamtin)
Lung cancer, ovarian cancer
GlaxoSmithKline, October 2007

Camptothecin (CPT)
Bark and stem of *Camptothec acuminata* (Happy Tree)

Docetaxel (DTX, DXL, Taxotere)
Breast, ovarian, lung, bladder, prostate, melanoma, esophageal
Sanofi-Aventis June 1998
(also sold as Docefrez by Sun Pharma Global and Zytax by Zydus.)

Paclitaxel (PTX or Taxol)
Bark of the Pacific yew tree
Taxus brevifolia

Vincristine (Oncovin)
Rosy Periwinkle *Catharanthus roseus*
Acute lymphocytic leukemia, acute myeloid
leukemia, Hodgkin's disease, neuroblastoma, and
small cell lung cancer
Eli Lilly and Company. July 1963

856

857 **Figure 5: Structures of semi-synthetic and natural products used in chemotherapy**

Figure 6: Activation of PI3K and Akt mediated through (a) mitogenic activation at the receptor tyrosine kinase (RTK) resulting in the (2) phosphorylation and binding of the PI3K intracellular unit (3) conversion of PIP₂ to PIP₃ and activation of the Ph subunit on Akt inducing the translocation to the cell membrane (4) phosphorylation and activation of Akt. Abbreviations: receptor tyrosine kinase (RTK), Phosphatidylinositol-4,5-bisphosphate 3-kinase (p85/p110kin), Phosphatidylinositol 4,5-bisphosphate (PIP), Pyruvate Dehydrogenase Kinase (PDK) and Integrin-linked kinase (ILK).

867 **11.0 Tables**

868 **Table 1: Proportion of Catechins present in Tea**

Fermentation	% Flavanols
Non-Fermented (Green)	8.0–14.4
Partially Fermented (Oolong)	4.14–4.92
Fermented (Black)	0.24–0.51

Table 2: EGCG's association as a chemopreventive with a range of cancers.

Cancer	Effect of EGCG	Source
Prostate	Cellular apoptosis	[109]
	Reduced risk	[36]
Lymphoma (mouse)	Cellular apoptosis	[109]
Keratinocytes	Cellular apoptosis	[109]
Bladder	Reduced Risk	[36]
Carcinoma	Chemoprevention	[4, 5]
	Prevent reoccurrence	[33]
	Fewer side effects	[126]
Colon	Chemoprevention	[57, 62, 127, 128]
	Cellular apoptosis	[10]
	Chemoprevention	[57]
	Inhibit tumour development (mouse/rat)	[57]
Lung	Controversial association with efficacy	[4]
	Apoptosis via triggering H ₂ O ₂ production (H661)	[118]
	Inhibit proliferation	[37]
Breast	Chemoprevention	[129]
	Inhibit proliferation	[37]
Lymphatic	Elevate ROS production during apoptosis	[36]
	Cellular apoptosis (mouse LY5178)	[109]
	Reduced adenocarcinoma incidence (Polyphenol E*)	[57]
	Reduced adenocarcinoma multiplicity (Polyphenol E*)	[57]
	Increase phosphorylation of cJun	[130, 131]
Epidermal	Increase phosphorylation of Erk1/2	[131]
	Increase PCNA	[131]
	G ₀ /G ₁ halt in A431 (not non-cancerous NHEK)	[11]
	Constitutively expression of NF-kB	[109]
	Apoptotic cell death	[109]
(Inflammation)	Protect against collagen-induced arthritis (GTP**)	[109]
Esophageal	Chemopreventive	[4, 57]
Stomach	Chemopreventive	[4, 57]
Intestine	Chemopreventive	[4, 57]

**polyphenol E is a concentrated catechin preparation; ** GTP refers to a combination of green tea proteins;*

Table 3: Eligibility criteria and common side effects for current PCa treatments.

	Drug	Brand Name (Manufacturer)	Adverse Reactions	Eligibility
GnRH Agonist / Antagonist	Goserelin	Zoladex (AstraZeneca Pharmaceuticals)	Hot flushes	locally advanced or metastatic hormone-sensitive prostate cancer
			Tumor flare	
			Hyperglycaemia	
			Hyperlipidemia	
			Hypercholesterolemia	
	Triptorelin	Decapeptyl (Ferring Pharmaceuticals)	Reduced libido	locally advanced or metastatic hormone-sensitive prostate cancer
			Hot flushes	
			Tumor flare	
			Hyperglycaemia	
			Hyperlipidemia	
Androgen Receptor Inhibitors	Degarelix	Firmagon (Ferring Pharmaceuticals)	Hypercholesterolaemia	Locally advanced or metastatic prostate cancer
			Reduced libido	
			Gynaecomastia	
			Fatigue	
			Constipation	
	Bicalutamide	Casodex (AstraZeneca Pharmaceuticals)	Nausea	locally advanced or metastatic CRPCa in combination with LHRH agonist
			Drowsiness	
			Constipation	
			Dizziness	
			Hot flushes	
Androgen Receptor Inhibitors	Cyproterone Acetate	Sandoz (Sandoz Pty Ltd)	Abdominal pain	Locally advanced inoperable prostate cancer in combination with radiation therapy Locally advanced or metastatic castrate- resistant prostate cancer in combination with LHRH agonist Short term prevention of tumor flare associated with the initiation of an LHRH agonist
			Fluid retention	
			Hepatotoxicity	
			Anorexia	
			Hot flushes	
Androgen Receptor Inhibitors	Abiraterone	Zytiga (Janssen Biotech)	Fatigue	Post-docetaxel CRPCa. secondary hormonal therapeutic. Must be in combination with prednisone or prednisolone, and no other chemotherapy
			Depression	
			Swelling	
			Bone weakening	
			Weight fluctuations	
Androgen Receptor Inhibitors	Enzalutimide	Xtandi (Astellas Pharma US)	Dry skin	Post-docetaxel CRPCa. Available for clinical trials secondary hormonal therapeutic. Cannot use with abiraterone.
			Hypertension	
			Fluid retention	
			Hypokalaemia	
			Vomiting and Diarrhoea	
Androgen Receptor Inhibitors			Hypertension	
			Anxiety	
			Fatigue	
			Seizures	

Flutamide	Eulexin (Schering-Plough)	Hot Flashes Urine Discolouration Loss of sexual interest Diarrhea Nausea Vomiting Enlargement of male breasts Skin sensitivity Impotence Rectal bleeding Nausea Vomiting Diarrhea Swelling	Locally advanced or metastatic castrate resistant prostate cancer. Used in combination with LHRH agonist throughout treatment. Short term prevention of tumour flare
Radium-223	Xofigo (Bayer)	Fatigue Fever Chills Nausea Vomiting Neutropenia Thrombocytopenia Oral Mucositis Diarrhea	Asymptomatic bone metastasis. Bone metastasis
Sipuleucel-T	Provenge (Dendreon Pharmaceuticals)	Skin rash Peripheral neuropathy Palmar-plantar Erythrodysaesthesia Arthralgia Ocular changes Fatigue Fluid retention Nausea Hair loss Mouth ulcers Neutropenia Thrombocytopenia Oral Mucositis Anorexia Arthralgia Fatigue Severe neutropenia Thrombocytopenia Anorexia Diarrhea Constipation Skin Rash Arthralgia Fatigue Peripheral neuropathy Peripheral neuropathy	Asymptomatic or minimally metastatic CRPCa
Docetaxel	Taxotere (Phyton Biotech)		Diagnosed CRPCa (standard treatment)
Mitoxantrone	Novantrone (Pfizer)		Diagnosed CRPCa
Cabazitaxel	Jevtana (Sanofi- Aventis)		Post-docetaxel CRPCa. Must be in combination with prednisone or prednisolone

Nausea and vomiting
Diarrhea

870	Note that this table is not fully comprehensive and the most current Australian data can be found at the EviQ
871	[132-140]

 872 |

Table 4: PI3K/Akt/mTOR pathway inhibitors			
Target		Name	Tradename
Dual PI3K/ mTOR		LY3023414	
		LY294002	
PI3K	Pan-class I Inhibitors	PX 866	Sonolisib
		BKM 120	Buparlisib
	p110 Isoform-specific	GSK 2636771	
	PI3K Inhibitors	AZD 8186	
AKT		GSK2141795	Uprosertib
	Akt	GDC-0068	Ipatasertib
		AZD5363	
	mTORC1 &	AZD 8055	
mTORC	mTORC2 Dual	INK 128	
	Inhibitor		
	MTORC1	Everolimus

Flavone

Flavonol

Flavanone

Flavan-3-ol

Flavanonol

Anthocyanidin

Chalcone

Aurone

Flavones

Apigenin

Lutelin

Flavonoids

Quercetin

Myricetin

Kaempferol

Flavan-3-ol (Catechins)

(-)-Epicatechin (EC)

(+)-Catechin (C)

(-)-Epigallocatechin (EGC)

(+)-Gallocatechin (GC)

(-)-Epicatechin gallate (ECG)

(-)-Catechin gallate (CG)

(-)-Epigallocatechin gallate (EGCG)

(-)-Gallocatechin gallate (GCG)

Topotecan (Hycamtin)
Lung cancer, ovarian cancer
GlaxoSmithKline, October 2007

Original
Source
→

Camptothecin (CPT)
Bark and stem of *Camptothec
acuminate* (Happy Tree)

Docetaxel (DTX, DXL, Taxotere)
Breast, ovarian, lung, bladder, prostate, melanoma, esophageal
Sanofi-Aventis June 1998
(also sold as Docefrez by Sun Pharma Global and Zytax by
Zydus.)

Original
Source
→

Paclitaxel (PTX or Taxol)
Bark of the Pacific yew tree
Taxus brevifolia

Vincristine (Oncovin)
Rosy Periwinkle *Catharanthus roseus*
Acute lymphocytic leukemia, acute myeloid
leukemia, Hodgkin's disease, neuroblastoma, and
small cell lung cancer
Eli Lilly and Company. July 1963

Tea Leaf

Withering

Panfrying or Steaming

Rolling and Shaping

Drying

Bruising

Short Fermentation

Panfrying and Drying

Bruising or Rolling

Full Fermentation

Firing and Dried

Components of Green Tea

Table 1: Proportion of Catechins present in Tea

Fermentation	% Flavanols
Non-Fermented (Green)	8.0–14.4
Partially Fermented (Oolong)	4.14–4.92
Fermented (Black)	0.24–0.51

Table 2: EGCG's association as a chemopreventive with a range of cancers.

Cancer	Effect of EGCG	Source
Prostate	Cellular apoptosis	[109]
	Reduced risk	[36]
Lymphoma (mouse)	Cellular apoptosis	[109]
Keratinocytes	Cellular apoptosis	[109]
Bladder	Reduced Risk	[36]
Carcinoma	Chemoprevention	[4, 5]
	Prevent reoccurrence	[33]
	Fewer side effects	[126]
Colon	Chemoprevention	[57, 62, 127, 128]
	Cellular apoptosis	[10]
	Chemoprevention	[57]
	Inhibit tumour development (mouse/rat)	[57]
Lung	Controversial association with efficacy	[4]
	Apoptosis via triggering H ₂ O ₂ production (H661)	[118]
	Inhibit proliferation	[37]
Breast	Chemoprevention	[129]
	Inhibit proliferation	[37]
Lymphatic	Elevate ROS production during apoptosis	[36]
	Cellular apoptosis (mouse LY5178)	[109]
	Reduced adenocarcinoma incidence (Polyphenol E*)	[57]
	Reduced adenocarcinoma multiplicity (Polyphenol E*)	[57]
	Increase phosphorylation of cJun	[130, 131]
Epidermal	Increase phosphorylation of Erk1/2	[131]
	Increase PCNA	[131]
	G ₀ /G ₁ halt in A431 (not non-cancerous NHEK)	[11]
	Constitutively expression of NF-kB	[109]
	Apoptotic cell death	[109]
(Inflammation)	Protect against collagen-induced arthritis (GTP**)	[109]
Esophageal	Chemopreventive	[4, 57]
Stomach	Chemopreventive	[4, 57]
Intestine	Chemopreventive	[4, 57]

**polyphenol E is a concentrated catechin preparation; ** GTP refers to a combination of green tea proteins;*

Table 3: Eligibility criteria and common side effects for current PCa treatments.

	Drug	Brand Name (Manufacturer)	Adverse Reactions	Eligibility
GnRH Agonist / Antagonist	Goserelin	Zoladex (AstraZeneca Pharmaceuticals)	Hot flushes Tumor flare Hyperglycaemia Hyperlipidemia Hypercholesterolemia Reduced libido	locally advanced or metastatic hormone-sensitive prostate cancer
			Hot flushes Tumor flare Hyperglycaemia Hyperlipidemia Hypercholesterolaemia Reduced libido	
	Triptorelin	Decapeptyl (Ferring Pharmaceuticals)	Depression Hot flushes Arthralgia Fatigue	locally advanced or metastatic hormone-sensitive prostate cancer
			Constipation Reduced libido Gynaecomastia Fatigue Constipation Nausea Drowsiness	
	Degarelix	Firmagon (Ferring Pharmaceuticals)	Constipation Reduced libido Gynaecomastia Fatigue Constipation Nausea Drowsiness	Locally advanced or metastatic prostate cancer
			Constipation Dizziness Hot flushes Abdominal pain Fluid retention Hepatotoxicity Anorexia	
	Bicalutamide	Casodex (AstraZeneca Pharmaceuticals)	Dizziness Hot flushes Abdominal pain Fluid retention Hepatotoxicity Anorexia	locally advanced or metastatic CRPCa in combination with LHRH agonist
			Hot flushes Fatigue Depression Swelling Bone weakening Weight fluctuations Dry skin	
	Cyproterone Acetate	Sandoz (Sandoz Pty Ltd)	Hypertension Fluid retention Hypokalaemia Vomiting and Diarrhoea	Locally advanced inoperable prostate cancer in combination with radiation therapy Locally advanced or metastatic castrate- resistant prostate cancer in combination with LHRH agonist Short term prevention of tumor flare associated with the initiation of an LHRH agonist
			Hypertension Fluid retention Hypokalaemia Vomiting and Diarrhoea	
Androgen Receptor Inhibitors	Abiraterone	Zytiga (Janssen Biotech)	Hypertension Fluid retention Hypokalaemia Vomiting and Diarrhoea	Post-docetaxel CRPCa. secondary hormonal therapeutic. Must be in combination with prednisone or prednisolone, and no other chemotherapy
	Enzalutamide	Xtandi (Astellas Pharma US)	Hypertension Anxiety Fatigue Seizures	Post-docetaxel CRPCa. Available for clinical trials secondary hormonal therapeutic. Cannot use with abiraterone.

Flutamide	Eulexin (Schering-Plough)	Hot Flashes Urine Discolouration Loss of sexual interest Diarrhea Nausea Vomiting Enlargement of male breasts Skin sensitivity Impotence Rectal bleeding	Locally advanced or metastatic castrate resistant prostate cancer. Used in combination with LHRH agonist throughout treatment.
Radium-223	Xofigo (Bayer)	Nausea Vomiting Diarrhea Swelling	Asymptomatic bone metastasis. Bone metastasis
Sipuleucel-T	Provenge (Dendreon Pharmaceuticals)	Fatigue Fever Chills Nausea Vomiting Neutropenia Thrombocytopenia Oral Mucositis	Asymptomatic or minimally metastatic CRPCa
Docetaxel	Taxotere (Phyton Biotech)	Diarrhea Skin rash Peripheral neuropathy Palmar-plantar Erythrodysesthesia Arthralgia Ocular changes Fatigue Fluid retention Nausea Hair loss Mouth ulcers	Diagnosed CRPCa (standard treatment)
Mitoxantrone	Novantrone (Pfizer)	Neutropenia Thrombocytopenia Oral Mucositis Anorexia Arthralgia Fatigue Severe neutropenia Thrombocytopenia Anorexia Diarrhea	Diagnosed CRPCa
Cabazitaxel	Jevtana (Sanofi-Aventis)	Constipation Skin Rash Arthralgia Fatigue Peripheral neuropathy Peripheral neuropathy Nausea and vomiting	Post-docetaxel CRPCa. Must be in combination with prednisone or prednisolone

Diarrhea

Note that this table is not fully comprehensive and the most current Australian data can be found at the EviQ [132-140]

Table 4: PI3K/Akt/mTOR pathway inhibitors			
Target		Name	Tradename
Dual PI3K/ mTOR		LY3023414	
		LY294002	
PI3K	Pan-class I Inhibitors	PX 866	Sonolisib
		BKM 120	Buparlisib
	p110 Isoform-specific PI3K Inhibitors	GSK 2636771	
		AZD 8186	
AKT		GSK2141795	Uprosertib
	Akt	GDC-0068	Ipatasertib
		AZD5363	
	mTORC1 & mTORC2 Dual Inhibitor	AZD 8055	
mTORC		INK 128	
	MTORC1	Everolimus	